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Cover: Juvenile moorhens, *Gallinula chloropus*, following and harassing an adult south-Italian asp viper, *Vipera aspis hugyi*, in Litorale di Ugento Regional Natural Park, southern Italy. Photograph by Roberto Gennaio.

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Off the Rails: Harassment of a South-Italian Asp Viper by Moorhens

Piero Carlino [1], Roberto Gennaio [2] and Olivier S. G. Pauwels [3]

The South-Italian Asp Viper *Vipera aspis hugyi* Schinz, 1833, is a venomous snake endemic to southern Italy and Sicily, and introduced to Montecristo Island (Masseti and Zuffi, 2011). Sometimes regarded as a distinct species, its natural history and precise geographic distribution are still relatively poorly documented. Recent publications reported the first cases of melanism (Di Nicola and Meier, 2013), of autophagy (Carlino and Pauwels, 2015), of predation on a centipede (Carlino et al., 2017), and of a patternless morph (Di Nicola and Faraone, 2020).

We report herein an unusual interaction between a South-Italian Asp Viper and birds in Litorale di Ugento Natural Park, Lecce Province, southern Italy. On 28 October 2023, while driving at 1:30 P.M. on a dirt road (39.885954°N, 18.123231°E) bordering the canal leading to Bacino Bianca water basin, in Torre San Giovanni, the attention of one of us (RG) was attracted by two juvenile Moorhens (Aves: Rallidae: *Gallinula chloropus* [Linnaeus, 1758]), which were deliberately following and pecking at an animal on the ground. It turned out to be a South-Italian Asp Viper, about 70 cm in total length, showing the typical adult dorsal color pattern of the species (Figure 1). The conditions of the observation were ideal, with a clear, sunny sky, and a temperature of 26°C. At this location the six-meter-wide road is bordered by a vegetation dominated by common reeds (Poaceae: *Phragmites australis*) and black rush (Cyperaceae: *Schoenus nigricans*). The viper crossed the road in a straight line, continuously followed by the rails which regularly pecked at it, until it disappeared into the vegetation on the other side. Although being harassed, the viper showed no signs of panic or aggressiveness, and continued its way.

This observation is the first documented interaction between the South-Italian Asp Viper and rallids. Moorhens feed mostly on plants, in a lesser proportion on small invertebrates, and only occasionally on small vertebrates such as small fish and tadpoles; extremely rarely they also feed on carrion (Lardjane-Hamiti et al., 2015). A live, adult viper is not a potential prey item for Moorhens, and hunting is definitely not what motivated the birds involved in our observation. Interactions between Moorhens and snakes are rarely reported in the literature, harassment of snakes is uncommon, and predation by rails on snakes is only exceptionally observed (Cutten et al., 2013). It is possible that the behavior of the rails was rather motivated by the fact that they regard the snake as a danger for themselves or as a potential predator of their offspring, and want to chase it out of their area of activity. Small birds are indeed known to be part of the diet of this viper (see Carlino et al. [2017] and references therein).

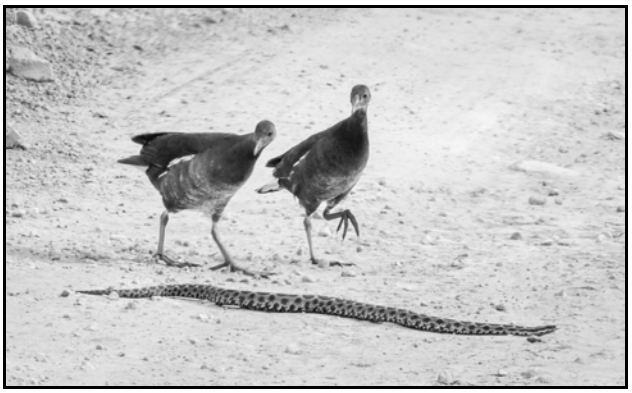

Figure 1. Juvenile Moorhens following and harassing an adult *Vipera aspis hugyi* in Litorale di Ugento Regional Natural Park, southern Italy. Photograph by R. Gennaio.

Acknowledgments

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Mexican Geographical Distribution Notes 10: New Distribution Records for Various Lizards (Squamata: Sauria) from Southern Jalisco, Mexico

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Introduction

Southern Jalisco is one of the herpetologically least studied regions of Jalisco, especially in the municipalities bordering Colima and Michoacán. During recent sampling of reptiles in the area, several interesting observations of lizards were made. We present eight new municipal records for species in the families Phrynosomatidae and Teiidae. These records were obtained during field work performed 03–04 June 2021 in the municipalities of Tecalitlán and Pihuamo, Jalisco. All specimens were identified by Jacobo Reyes-Velasco. Digital photographs of all individuals reported herein were deposited at the *Colección Digital de Vertebrados de la Facultad de Estudios Superiores Zaragoza*, UNAM (MZFZ-IMG). The coordinates and elevations of each location were taken by GPS device using the WGS 84 cartographic datum. Distances reported between locations are given as straight-line kilometers.

Distribution Records

Teiidae

Holcosus sinister (Smith y Laufe, 1946).
MEXICO: JALISCO: Municipio de Pihuamo: 10 km N of La Estrella (19.278761°N; 103.503978°W; 688 m elev; Figure 1). 03 June 2021. MZFZ-IMG 435. First municipal record, this record extends the known range approximately 13 km SE from the closest known locality at Quesería, Colima (Smith and Laufe, 1946). This individual was photographed at 12:00 P.M. on the edge of tropical deciduous forest and an agricultural field.

Aspidoscelis communis (Cope, 1878).
MEXICO: JALISCO: Municipio de Pihuamo: 10 km N of La Estrella (19.274364°N; 103.501344°W; 680 m elev.; Figure 2). 03 June 2021. MZFZ-IMG 433. First municipal record, this record extends the known range approximately 12 km SE from the closest known locality at Quesería, Colima (Zweifel, 1959).

Figure 1. *Holcosus sinister*, Municipio de Pihuamo.

Figure 2. *Aspidoscelis communis*, Municipio de Pihuamo.

This individual was photographed during the morning on the edge between tropical deciduous forest and an agricultural field.

Aspidoscelis costatus (Cope, 1878).
MEXICO: JALISCO: Municipio de Tecalitlán: 1.8 km W of Tecalitlán
(19.465617°N; 103.323842°W; 1134 m elev.; Figure 3). 04 June 2021. MZFZ-IMG 425-427. First municipal record, and

Figure 3. *Aspidoscelis costatus*, Municipio de Tecalitlán.

1. Biodiversa A.C., Avenida de la Ribera #203, C.P. 45900, Chapala, Jalisco, Mexico.

2. Biencom Real Estate, Carretera Chapala–Jocotepec #57-1, C.P. 45920, Ajijic, Jalisco, Mexico.

3. Herp.mx A.C., C.P. 28989, Villa de Álvarez, Colima, Mexico.

Figure 4. *Aspidoscelis lineattissimus*, Municipio de Pihuamo.

extends the known range approximately 11 km SE from Tuxpan, Jalisco (Zweifel, 1959). This individual was observed active during the day in an agricultural field.

Aspidoscelis lineattissimus (Cope, 1878).
MEXICO: JALISCO: Municipio de Pihuamo: 10 km N of La Estrella (19.276661°N; 103.504467°W; 662 m elev.; Figure 4). 03 June 2021. MZFZ-IMG 434. First municipal record and extends the known range approximately 21 km NE from "8 km E of Colima" (Duellman and Wellman, 1960). This individual was observed at noon on the edge of tropical deciduous forest.

Phrynosomatidae
Sceloporus melanorhinus (Bocourt, 1876).
MEXICO: JALISCO: Municipio de Tecalitlán: 1.8 km W of Tecalitlán (19.463392°N; 103.322464°W; 1136 m elev.; Figure 5). 04 June 2021. IMG-MZFZ 428. First municipal record, and extends the known range of this species approximately 27 km ENE from Quesería, Colima (Smith, 1939). This lizard was found on a wall in an abandoned building in an agricultural field.

MEXICO: JALISCO: Municipio de Pihuamo: 9 km N of La Estrella (19.274822°N; 103.496797°W; 635 m elev.). 03 June 2021. IMG-MZFZ 429. First municipal record, and extends the known range of this species 14 km SE from the closest published locality at Quesería, Colima (Smith, 1939). This individual was photographed on a tree trunk in tropical deciduous forest.

Figure 5. *Sceloporus melanorhinus*, Municipio de Tecalitlán.

Sceloporus pyrocephalus (Cope, 1864).
MEXICO: JALISCO: Municipio de Pihuamo: 10 km N of La Estrella (19.273756°N; -103.501356°W; 671 m elev., Figure 6). 03 June 2021. IMG-MZFZ 430. This is the first municipal record and extends the known range of this species 18 km E of the closest published locality at "2 miles E of Colima" (Smith, 1939). Several individuals of this species were observed on large rocks at the edge of a tropical deciduous forest.

Sceloporus utiformis (Cope, 1864).
MEXICO: JALISCO: Municipio de Pihuamo: 5.5 km N of La Estrella (19.203983°N; 103.504964°W; 596 m. elev., Figure 7).

Figure 6. *Sceloporus pyrocephalus*, Municipio de Tecalitlán.

Figure 7. *Sceloporus utiformis*, Municipio de Pihuamo.

03 June 2021. IMG-MZFZ 431. First municipal record, and extends known range approximately 21 km E from the closest known localities in Colima (Smith, 1939). This species was common and numerous individuals were found active on on the forest floor in tropical deciduous forest.

Urosaurus bicarinatus (Duméril, 1856).
MEXICO: JALISCO: Municipio de Pihuamo: 10 km N of La Estrella (19.274722°N; 103.498506°W; 655 m elev.; Figure 8). 03 June 2021. IMG-MZFZ 432. This record representes a first municipal record and extends the known range for this species 20 km E of the closest known locality in Colima (Mittleman, 1942). One individual was observed thermoregulating on the main trunk of an unidentified species of tree in tropical deciduous forest.

Figure 8. *Urosaurus bicarinatus*, Municipio de Pihuamo.

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Notes on Reproduction of Barking Treefrogs, *Dryophytes gratiosus* (Anura: Hylidae), from North Carolina

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Abstract

I conducted a histological examination of gonads from 23 *Dryophytes gratiosus* adults from North Carolina consisting of 11 males and 12 females. Males contained sperm from all months examined: February to October. The two smallest mature males (sperm in lumina of seminiferous tubules) measured 55 mm SVL and were from April and August. Females in spawning condition were present in all months: April to October. The smallest mature female (spawning condition) measured 50 mm SVL and was from June. Three (25%) of my sample of 12 females contained atretic follicles. One female from May and one from July contained postovulatory follicles (evidence of recent spawning). The May *D. gratiosus* postovulatory female contained numerous concurrent vitellogenic follicles and would have likely had sufficient time to spawn a second time in the same reproductive period. However, it is uncertain if the few vitellogenic follicles in the July *D. gratiosus* postovulatory female would have completed yolk deposition and spawned in the same reproductive period. My findings of males exhibiting spermiogenesis and gravid females from September and October confirms published reports that some *D. gratiosus* reproduction occurs in autumn.

Dryophytes gratiosus (LeConte, 1856) occurs in the southeastern United States (eastern Virginia to eastern Louisiana) with isolated records from Kentucky, New Jersey, Delaware, Tennessee and Alabama (Frost, 2023). *Dryophytes gratiosus* breeds in a wide variety of shallow wetlands; adults migrate from arboreal and terrestrial retreats to breeding sites (Mitchell, 2005). For breeding to occur, habitats should contain temporary wet areas (Dodd, 2023). *Dryophytes gratiosus* usually stay in trees or shrubs or burrow into damp sand when not involved in reproductive activities; they are not often seen except after heavy rains (Green et al., 2013). The biology of *D. gratiosus* is summarized in Caldwell (1982). In the current paper I present data on the *D. gratiosus* reproductive cycle from a histological examination of gonadal material from North Carolina. Utilization of museum collections for obtaining reproductive data avoids removing additional animals from the wild.

A sample of 23 *D. gratiosus* from North Carolina collected 1940 to 1988 consisting of 11 adult males (mean SVL = 60.4 mm ± 4.4 SD, range = 55–68 mm) and 12 adult females (mean SVL = 56.7 mm ± 4.3 SD, range = 50–65 mm) was examined from the herpetology collection of the North Carolina State Museum of Natural Sciences (NCSM), Raleigh, North Carolina, USA (Appendix). An unpaired *t*-test was used to test for differences between adult male and female SVLs.

A small incision was made in the lower part of the abdomen; the left testis was removed from males and a piece of the left ovary from females. Gonads were embedded in paraffin, sections were cut at 5 μm and stained with Harris hematoxylin followed by eosin counterstain (Presnell and Schreibman, 1997). Histology slides were deposited at NCSM.

There was no significant size difference between SVLs of adult males versus adult females of *D. gratiosus* ($t = 2.1$, df = 21, $P = 0.05$). The testicular morphology of *D. gratiosus* is similar to that of other anurans as described in Ogielska and Bartmañska (2009a). Within the seminiferous tubules, spermatogenesis occurs in cysts that stay closed until the late spermatid stage is reached; cysts then open and differentiating sperm reach the lumina of the seminiferous tubules (Ogielska and Bartmañska, 2009a). All 11 *D. gratiosus* males were undergoing sperm formation (= spermiogenesis) in which clusters of sperm filled the seminiferous tubules. A ring of germinal cysts was located on the inner periphery of each seminiferous tubule. By month, numbers of *D. gratiosus* males exhibiting spermiogenesis were: February (N = 1), April (N = 1), May (N = 1), June (N = 2), July (N = 1), August (N = 1), September (N = 2), October (N = 2). The two smallest mature males (sperm in lumina of seminiferous tubules) measured 55 mm SVL and were from April (NCSM 102155) and October (NCSM 98856). Wright and Wright (1949) reported adult *D. gratiosus* males ranged from 49 to 68 mm in body size.

The ovaries of *D. gratiosus* are typical of other anurans in consisting of paired organs located on the ventral sides of the kidneys; in adults they are filled with diplotene oocytes in various stages of development (Ogielska and Bartmañska, 2009b). Mature oocytes are filled with yolk droplets; the layer of surrounding follicular cells is thinly stretched. Two stages were present in the spawning cycle (Table 1): (1) "Postovulatory follicles; not in spawning condition" (2) "Ready to spawn

Table 1. Two monthly stages in the spawning cycle of 12 adult female *Dryophytes gratiosus* from North Carolina.

Month	*n*	Postovulatory follicles; not in spawning condition	Ready to spawn condition
April	2	0	2
May	4	1	3
June	2	0	2
July	2	1	1
September	1	0	1
October	1	0	1

Table 2. Periods of reproduction for *Dryophytes gratiosus* from different localities.

Locality	Breeding Period	Source
Alabama	call February to August	Guyer and Bailey, 2023
Carolinas and Virginia	late spring and summer	Beane et al., 2010
Florida	call March to August	Carr, 1940
Florida	February to October	Krysko et al., 2019
Florida (west central)	April to August	Delis, 2001
Georgia	March to August	Jensen et al., 2008
Louisiana	late March to mid-August	Boundy and Carr, 2017
No specific locality	March to August	Wright and Wright, 1949
North Carolina	April to September	Dorcas et al., 2007
Southeast	March or April to late summer	Dorcas and Gibbons, 2008
Tennessee	late spring or summer	Niemiller and Reynolds, 2011

condition" in which mature oocytes predominate. The smallest mature *D. gratiosus* female (ready to spawn condition) measured 50 mm SVL (NCSM 62310) and was from June. Wright and Wright (1949) reported adult *D. gratiosus* females ranged from 50 to 68 mm in body size.

Postovulatory follicles form when the ruptured follicle collapses after ovulation; the follicle lumen disappears and proliferating granulosa cells are surrounded by a fibrous capsule (Redshaw, 1972). Postovulatory follicles are short-lived in most anuran species and are resorbed after a few weeks (Redshaw, 1972). The presence of postovulatory follicles (remnant of previous spawning) and vitellogenic follicles (undergoing yolk deposition) in the same ovary suggests a second spawning will occur in the same reproductive period. There were two cases of postovulatory follicles in my current sample of *D. gratiosus* (Table 1). The first was in (NCSM 39598) from May (early in the reproductive season) contained concurrent postovulatory follicles and numerous vitellogenic follicles which would have likely spawned later the same year. The second case from July (NCSM 102033) exhibited a different condition. Postovulatory follicles were apparent but only a few vitellogenic follicles were present. Considering it was late in the reproductive season (July), I doubt if this female would have spawned a second time during the same reproductive season.

Atretic follicles were noted in the ovaries of 3/12 (25%) of my *D. gratiosus* female sample (Table 1). In early atresia the granulosa layer is slightly enlarged and contains ingested yolk granules. In late atresia the oocytes of these females are replaced by brownish, vacuolated granulosa cells which invade the lumen of the oocyte, or by solid black pigment-containing cells. Atresia is a widespread process occurring in the ovaries of all vertebrates (Uribe Aranzábal, 2009). It is common in the amphibian ovary (Saidapur, 1978) and is the spontaneous digestion of a diplotene oocyte by its own hypertrophied and phagocytic granulosa cells which invade the follicle and eventually degenerate after accumulating dark pigment (Ogielska and Bartmañska, 2009b). See Saidapur and Nadkarni (1973) and Ogielska et al. (2010) for detailed descriptions of follicular atresia in the frog ovary. Atresia plays an important role in fecundity by influencing numbers of ovulated oocytes (Uribe Aranzábal, 2011). The causes of follicular atresia in non-mammalian vertebrates are not fully understood although it is associated with captivity, food availability, crowding and irradiation (Saidapur, 1978). In amphibians, adverse environmental conditions such as starvation and suboptimal lighting may cause atresia of vitellogenic oocytes (Jørgensen, 1992). Incidences of follicular atresia increase late in the reproductive period (Saidapur, 1978). Saved energy will be presumably utilized during a subsequent reproduction.

Times of breeding for *D. gratiosus* throughout its range are shown in Table 2. My findings of males exhibiting spermiogenesis and gravid females (Table 1) from September and October confirms published reports (Table 2) that some *D. gratiosus* reproduction occurs in autumn (Dorcas et al., 2007; Krysko et al., 2019). The May *D. gratiosus* female with postovulatory follicles and concurrent vitellogenic follicles (NCSM 39598) would have likely had sufficient time to spawn a second time. This indicates the possibility that some *D. gratiosus* may spawn a second time in the same reproductive period which apparently occurs in some other North American hylid frogs (see Goldberg, 2023).

Acknowledgments

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Appendix

Twenty-three *D. gratiosus* from North Carolina examined by county from the herpetology collection of the North Carolina State Museum (NCSM), Raleigh. North Carolina. **Bladen:** NCSM: 49980, 98942, 100606, 100692; **Carteret:** NCSM: 9641; **Moore**: NCSM**:** 62310, 63292, 68822; **New Hanover:** NCSM: 84734; **Onslow**: NCSM: 39598; **Pender**: NCSM**:** 16143; **Richmond**: NCSM: 102032, 102033; **Sampson**: NCSM**:** 60111, 96889, 97032, 98777, 98791, 98792, 98856; **Scotland**: NCSM: 38815, 49983, 102155.

Herpetological Sculpture at the John Ball Zoo, Grand Rapids, Michigan

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Madison, WI 53707-7921
dreux.watermolen@wisconsin.gov

Located in a 103-acre park and occupying nearly 31 acres, the John Ball Zoo remains one of Michigan's most attended cultural facilities. The zoo traces its roots to 1884 when Mr. John Ball died and left 40 acres of land to the City of Grand Rapids for public use. By 1891, references to animals being kept at "Ball 40" appeared, and a local newspaper reported on a small menagerie with various kinds of owls, two hawks, a crow, an eagle, raccoons, a woodchuck, fox squirrels, and rabbits. In November 1891, two city leaders donated money from their salaries to purchase a pair of deer to start a herd to populate the area's hillside. Today, a 501(c)(3) nonprofit corporation operates an Association of Zoos and Aquariums (AZA) accredited zoo at the site with more than 220 species and over 2,400 individual animals, including a variety of amphibians and reptiles. The zoo also features several herpetological sculptures that may interest CHS members.

Visitors to the zoo's meerkat exhibit will encounter a small bronze sculpture of a king cobra (*Ophiophagus hannah*; Figure 1). Rising about a foot, the sculpture is nestled in a rock alcove along the exhibit's perimeter. The meerkat exhibit is the first of the zoo's exhibits created in adherence with the Living Building Challenge, a program centered on creating sustainable buildings through sustainable processes.

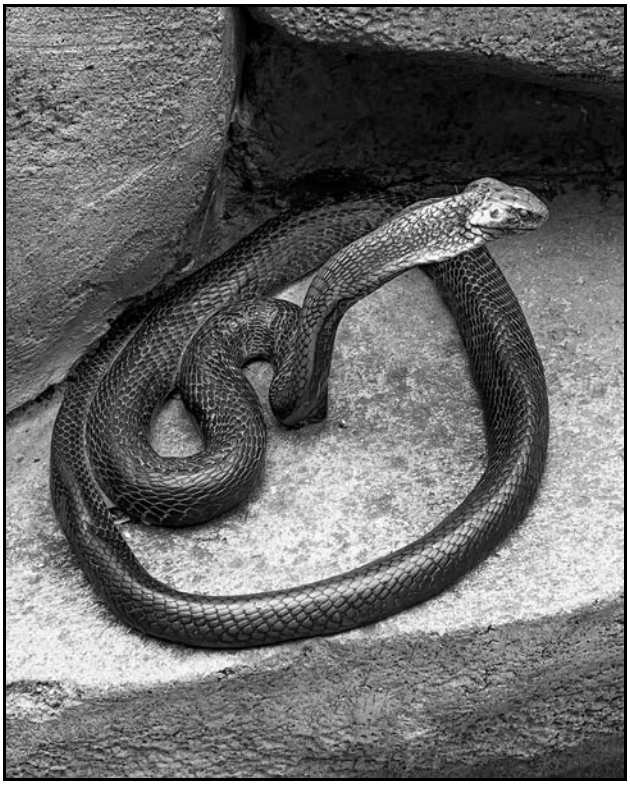

Figure 1. A bronze sculpture of a king cobra near the John Ball Zoo's meerkat exhibit. Photograph by Dreux J. Watermolen.

Ten sculptures resembling geckos (Gekkonidae) and tree-frogs (Hylidae) can be found "crawling" on the outside wall of the zoo's Monkey Island Café (Figure 2). Reminiscent of the popular chia pets, these sculptures provide a substrate for various plants. The zoo's horticulture staff regularly maintains these sculptures.

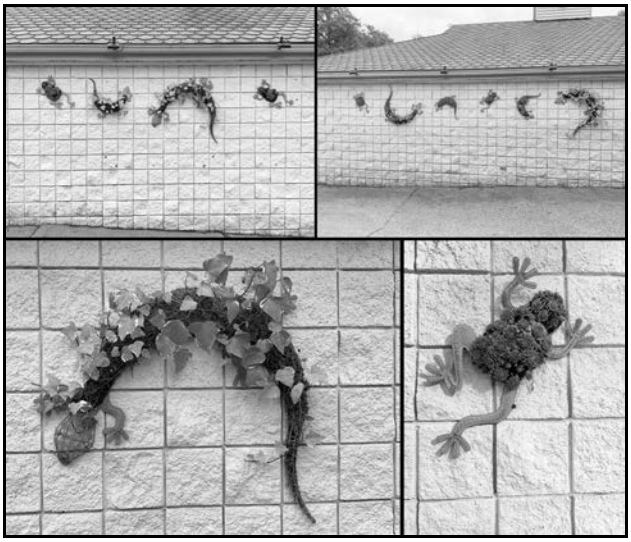

Figure 2. Wall-hanging gecko and frog sculptures adorn the side of John Ball Zoo's Monkey Island Café. **Top:** All ten sculptures. **Bottom:** Close-ups of one of the geckos and one of the frogs. Photographs by Dreux J. Watermolen.

A short distance from Monkey Island Café, visitors can enter the Natural Treasures building which houses various turtles, lizards and snakes (Figure 3), along with small primates, armadillos, and an assortment of invertebrates.

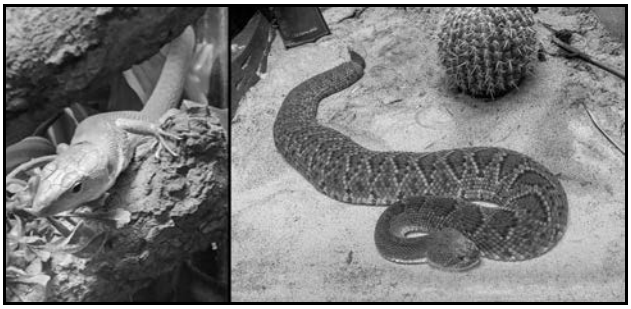

Figure 3. A green keel-bellied lizard (*Gastropholis prasina*) and a Mexican west coast rattlesnake (*Crotalus basiliscus*), two of the species on exhibit in John Ball Zoo's Natural Treasures building.

The zoo's only living crocodilians are a pair of Cuvier's dwarf caimans (*Paleosuchus palpebrosus*) housed in the Natural Treasures building, but visitors may encounter an interesting alligator-like sculpture near the Budgie Aviary. A floating head

covered in aquatic vegetation gives the impression of a submerged alligator peering above the water's surface (Figure 4), although zoo staff indicate it is actually a hippopotamus. The head floats in a pond at the base of a series of small waterfalls.

Figure 4. An "alligator" head floats in the pool at the base of John Ball Zoo's four-tier waterfall. Photograph by Dreux J. Watermolen.

During the 2022 visitor season John Ball Zoo hosted a collection of 16 intricately designed sea life sculptures made entirely of marine debris collected from the beaches of Oregon. The colorful sculptures, located throughout the zoo, graphically illustrated the tragedy of plastic pollution in our oceans and waterways in an effort to inspire active engagement in ocean conservation. The Washed Ashore Traveling Exhibit included a sculpture of *Natasha the Sea Turtle* (Figure 5).

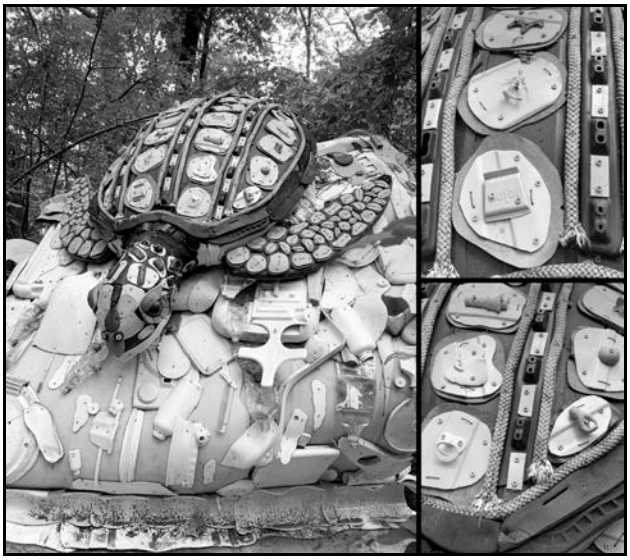

Figure 5. *Natasha the Sea Turtle* sculpture comprised of marine debris. **Left:** Statute in situ. **Right:** Close-ups of some of the debris used in the construction of the sculpture. Photographs by Dreux J. Watermolen.

For eight weeks in 2023, the zoo hosted a large Asian lantern festival installed by Tianyu Arts & Culture, Inc. The display featured handcrafted silk lanterns that illuminated the zoo and highlighted the intersecting story of wildlife and Asian culture. The display, which included several herp sculptures (Figure 6), attracted over 100,000 visitors and was so popular with the public that the zoo is planning a future installment.

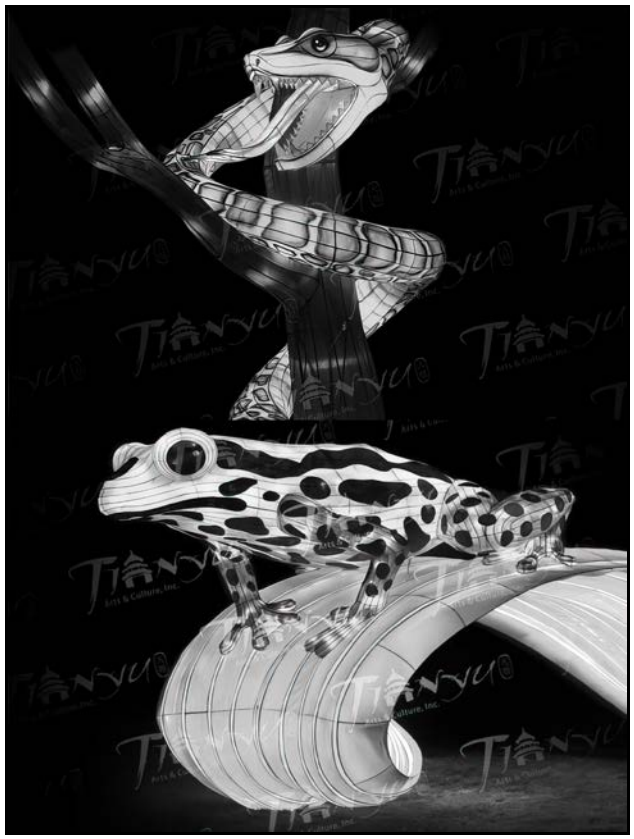

Figure 6. A display of silk lanterns by Tianyu Arts & Culture, Inc. at the John Ball Zoo included several amphibian and reptile themed lanterns like this boa and treefrog. Photos © taken from Tianyu Arts & Culture website <https://tianyuculture.us/new-theme-new-mission/>.

Visitors will be a little less likely to encounter the zoo's boa constrictor sculpture (Figure 7). This lifelike, papier mâché construction is tucked away in the ballroom in the upper level of the zoo administration building.

Figure 7. This papier mâché boa constrictor resides in the ballroom in the upper level of John Ball Zoo's administration building. Photograph by Dreux J. Watermolen.

John Ball Zoo hosts over 600,000 visitors annually and reaches over 800,000 people with its conservation messages. The zoo participates in 49 species survival plans and 39 taxon advisory groups. Charity Navigator gives John Ball Zoo a "4 Stars/Exceptional" rating (only 10% of AZA-accredited zoos and aquariums reach this high standard). The zoo is open daily. To find out more about John Ball Zoo, visit www.jbzoo.org/.

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Minutes of the CHS Board Meeting, November 14, 2023

A virtual meeting of the CHS board of directors via Zoom conference video/call was called to order at 7:52 P.M. Board member Gabrielle Evans was absent. No nonmembers of the board were present. Minutes of the September 12 board meeting were read and accepted.

Officers' reports

Treasurer: Rich Crowley went over the October financial report. Rich plans to send out information about how to donate to the Society on Giving Tuesday..

Media secretary: No immediate update. However, Stephanie is overwhelmed at the moment, especially with Facebook. She has asked if each board member could help out by taking a day and checking in on our Facebook page and removing any inappropriate content.

Membership secretary: Mike Dloogatch read the list of those whose memberships have expired Currently membership is just under 300. If we fall below 200 members it will become prohibitively expensive to mail the Bulletins, as we will no longer qualify for the nonprofit periodicals postage rate.

Sergeant-at-arms: Tom Mikosz reported that 20 people attended the October 15 meeting in person, and three viewed online.

Committee reports

Junior Herpers: Caitlin Monesmith will not be present for the December 17 meeting. We will roll it into the general meeting / holiday party and include a Show & Tell.

New Business

The December 17 general meeting / holiday party will be in-person only.

The meeting adjourned at 8:28 P.M.

Respectfully submitted by recording secretary Gail Oomens

Motions completed via email

October 10, 2023: Mike Dloogatch moved to remove John Archer as the corporation registered agent of the Chicago Herpetological Society to the Illinois Secretary of State, and replace him with Rich Crowley. Tom Mikosz seconded.
Yea votes—Mike, Tom, Amelia, Margaret-Ann, Gail, Gabby.
Motion Passed.

October 10, 2023: Mike Dloogatch moved to appoint Caitlin Monesmith as member-at-large of the CHS board, as provided for in our by-laws. Tom Mikosz seconded.
Yea votes—Mike, Tom, Amelia, Margaret-Ann, Gail, Kyle, Gabby.
Motion Passed.

November 15, 2023: Amelia Pollock moved to close the existing Chicago Herpetological Society money market account and checking account and establish new accounts with only Rich Crowley and Michael Dloogatch as signers on the new accounts. Mike Dloogatch seconded.
Yea Votes—Mike, Amelia, Margaret-Ann, Kyle, Gail, Gabby.
Motion Passed.

UPCOMING MEETINGS

Monthly meetings of the Chicago Herpetological Society begin at 2:00 P.M. on the third Sunday of each month. Please try to join us *in person* at the Notebaert Nature Museum, 2430 N. Cannon Drive, Chicago. **The December 17 meeting will be a holiday party**. The CHS will provide soft drinks and snacks. If you would like to bring something edible to share with the group, you are invited to do so. If you would like to bring an animal to show off to the group, you are encouraged to do that as well. This will be a chance to socialize and get to know your fellow members a little better.

The program for the January 21, 2024, meeting has not yet been confirmed.

Please check the CHS website or Facebook page each month for information on the program. Information about attending a Zoom webinar can be found here:
<https://support.zoom.us/hc/en-us/articles/115004954946-Joining-and-participating-in-a-webinar-attendee->

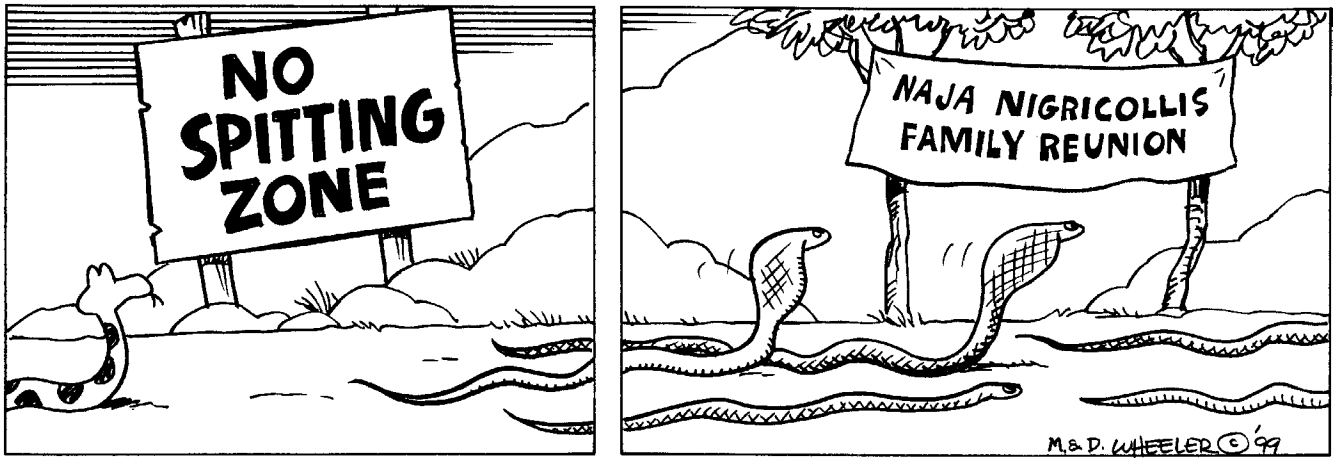
Board of Directors Meeting

Are you interested in how the decisions are made that determine how the Chicago Herpetological Society runs? And would you like to have input into those decisions? The next board meeting will be held online. If you wish to take part, please email: rcrowley@chicagoherp.org.

NEW CHS MEMBERS THIS MONTH

Jonah Bennett

THE ADVENTURES OF SPOT



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